

# Package ‘distory’

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**Type** Package

**Title** Distance Between Phylogenetic Histories

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**Maintainer** ORPHANED

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**Description**

The 'distory' package provides functions for computing geodesic distances between phylogenetic trees, as well as functions which use this distance. Methods for computing Gromov delta-hyperbolicity, Markov Chain Monte Carlo routines in tree space, and per-position leverage for DNA sequences are included.

**Details**

A description of the algorithm used for the distance computation can be found in `help(dist.multiPhylo)`.

**Author(s)**

John Chakerian <chakj@stanford.edu> and Susan Holmes <susan@stat.stanford.edu>

**References**

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

Billera, L. J., Holmes, S. P., and Vogtmann, K. (2001) Geometry of the space of phylogenetic trees. *Adv. Appl. Math.*, \*27\*, 733-767.

Megan Owen, J. Scott Provan, "A Fast Algorithm for Computing Geodesic Distances in Tree Space," IEEE/ACM Transactions on Computational Biology and Bioinformatics, 14 Jan. 2010.

**Examples**

```
data(woodmouse)
otree <- root(nj(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
breps = 250

trees <- boot.phylo(otree, woodmouse, B=breps, function(x)
  root(nj(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)

combined.trees <- c(list(otree), trees$trees)
tree.dists <- dist.multiPhylo(combined.trees)

mdres <- cmdscale(tree.dists, k=breps, add=TRUE)
plot(mdres$points[,1], mdres$points[,2], col = c("red", rep("black", breps)))
text(mdres$points[,1], mdres$points[,2], labels=1:(breps+1), cex=0.7, adj=c(0,2))
```

---

`bethe.tree`*Bethe Tree*

---

**Description**

Generates a Bethe tree with given tips, inner edge lengths, and outgroup.

**Usage**

```
bethe.tree(tips, level.lengths = NULL, outgroup="0", outgroup.dist=1)
```

**Arguments**

<code>tips</code>	A list of tip names as a character vector. Should be a power of 2. All tip names must be distinct.
<code>level.lengths</code>	Edge lengths for each level, counted from the bottom up. NULL means a default of 1. If the vector isn't long enough, the last value will be repeated as necessary.
<code>outgroup</code>	The tip label for the outgroup.
<code>outgroup.dist</code>	The distance of the outgroup from the root.

**Details**

Generates a Bethe tree with specified internal edge lengths.

**Value**

A class of type `phylo` representing the tree.

**Author(s)**

John Chakerian <chakj@stanford.edu>

**References**

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

**See Also**

[dist.multiPhylo](#)

**Examples**

```
plot(bethe.tree( as.character(1:16), 1:4, "17", 14))
```

---

`bin.multiPhylo`*Bin Trees*

---

**Description**

Bins trees according to branching topology.

**Usage**

```
bin.multiPhylo(treelist)
```

**Arguments**

`treelist` A list of trees that can be passed to `dist.phylo` (see the help for `dist.phylo` for acceptable formats)

**Details**

Bins trees according to branching topology. Two trees are considered to have the same topology if the same set of partitions of tips are produced by the edges, which corresponds to the same branching up to rearrangement of tips.

**Value**

Returns a numeric vector of bin ids. Bin ids are assigned in order of the first tree in that bin, that is, the first `k` unique trees in the list passed will be assigned bins 1..`k` in order of appearance.

**Author(s)**

John Chakerian <chakj@stanford.edu>

**References**

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

**See Also**

[dist.multiPhylo](#)

**Examples**

```
data(woodmouse)
otree <- root(fastme.ols(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
breps = 500

trees <- boot.phylo(otree, woodmouse, B=breps, function(x)
  root(fastme.ols(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)
```

```
combined.trees <- c(list(otree), trees$trees)

bin.multiPhylo(combined.trees)
```

---

dist.multiPhylo      *Geodesic Distance Between Phylogenetic Trees*

---

### Description

Computes the geodesic distance of a list of phylogenetic trees using a polynomial algorithm.

### Usage

```
dist.multiPhylo(x, method="geodesic", force.multi2di = FALSE, outgroup = NULL,
  convert.multifurcating = FALSE, use.random.resolution = FALSE, scale =
  NULL, verbose = FALSE)
```

### Arguments

x	A list of ape trees (class 'phylo'). The list does not have to be of class 'multiPhylo'. The function will also accept a list of strings of trees in Newick format, or a single string with trees in Newick format separated by semicolons. All the trees must have the same tip labels.
method	Determines which distance method is used. Options are 'geodesic' for the tree space geodesic distance, or 'edgeset' for the number of edges (defined by splits of tips) that are different.
force.multi2di	Force conversion of every tree to strict bifurcating through the ape function 'multi2di', using the use.random.resolution as its parameter. This option should not be used in conjunction with specification of an outgroup.
outgroup	Specifies an outgroup to root each tree with respect to. This calls the ape function 'root' on every tree in the list.
convert.multifurcating	Setting this option will check every tree for multifurcations using the ape function 'is.binary.tree' - if it returns FALSE, the ape function 'multi2di' will be called on it. Note that this does not ensure a tree is strictly binary, since ape considers an unrooted tree binary even if the root node is trifurcating. This option can be used in conjunction with specification of an outgroup.
use.random.resolution	Specifies the parameter to 'multi2di' if needed.
scale	Specifies a scale to make all trees uniformly scaled (that is, the sum of all edges will be uniform)scale to make all trees uniformly scaled (that is, the sum of all edge lengths will be uniform). The parameter can either be a tree of class phylo or a numeric value for the sum of all edge lengths.
verbose	Turns on incremental status updates and more warnings. Helpful for large computations.

## Details

This function computes the geodesic distance according to Billera et. al. using an algorithm based off of the polynomial time algorithm of Owen and Provan. Since it corresponds to a formal definition of tree-space as a space of strictly binary trees, no multifurcations are allowed, including on the root node. In addition, negative and 0-lengthed edges are clamped to a very small value (DBL\_MIN) for technical reasons.

The Newick parser supports only a subset of the Newick format. In particular, it does not at the moment allow for internal node labels, only weights. Weights will be automatically set to 1 if not specified. It may be necessary to clean data in ape to make the trees conform to this.

## Value

Returns a distance matrix of class 'dist' representing the pairwise geodesic distances between all input trees. Keep in mind this distance matrix is not Euclidean. N/A values are provided in the case of an error in determining the distance.

## Author(s)

John Chakerian <chakj@stanford.edu>

## References

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

Billera, L. J., Holmes, S. P., and Vogtmann, K. (2001) Geometry of the space of phylogenetic trees. *Adv. Appl. Math.*, \*27\*, 733-767.

Megan Owen, J. Scott Provan, "A Fast Algorithm for Computing Geodesic Distances in Tree Space," IEEE/ACM Transactions on Computational Biology and Bioinformatics, 14 Jan. 2010.

## See Also

[dist.dna boot.phylo cmdscale](#)

## Examples

```
data(woodmouse)
otree <- root(nj(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
breps = 250

trees <- boot.phylo(otree, woodmouse, B=breps, function(x)
  root(nj(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)

combined.trees <- c(list(otree), trees$trees)
tree.dists <- dist.multiPhylo(combined.trees)

mdres <- cmdscale(tree.dists, k=breps, add=TRUE)
plot(mdres$points[,1], mdres$points[,2], col = c("red", rep("black", breps)))
text(mdres$points[,1], mdres$points[,2], ,labels=1:(breps+1), cex=0.7, adj=c(0,2))
```

---

gromov.hyperbolicity *Gromov Hyperbolicity Constant*

---

### Description

Computes the Gromov Hyperbolicity Constant of a distance matrix.

### Usage

```
gromov.hyperbolicity(d, deltas = FALSE, scale = NA)
```

### Arguments

d	A distance matrix of type <code>dist</code> or <code>matrix</code> , or anything that can be coerced into <code>dist</code> by <code>as.dist</code> . Must have at least 4 points.
deltas	A logical value specifying whether to return the vector of delta values. Default is <code>FALSE</code> .
scale	Specifies a scaling method for each delta. Default is no scaling ( <code>NA</code> or <code>"none"</code> ). Available methods are <code>"max"</code> which scales deltas by the max of the sums computed, and <code>"perimeter"</code> for the largest perimeter of the four points.

### Details

This computes a constant that represents the relaxation of a 4-point condition for delta-hyperbolicity. See (Gromov 1987) for details.

### Value

The Gromov hyperbolicity constant of the given distance matrix.

### Author(s)

John Chakerian <chakj@stanford.edu>

### References

M. Gromov. Hyperbolic groups. In *Essays in Group Theory*, pages 73-263. Springer, New York, 1987.

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

### See Also

[dist.multiPhylo](#)

**Examples**

```
# scale final delta by max distance
points <- cbind(runif(100), runif(100))
d <- dist(points)
gromov.hyperbolicity(d)/max(d)

# scale each delta by max distance for the 4 points
points <- cbind(runif(100), runif(100))
d <- dist(points)
gromov.hyperbolicity(d, scale="max")

# scale each delta by the max perimeter for the 4 points
points <- cbind(runif(100), runif(100))
d <- dist(points)
gromov.hyperbolicity(d, scale="max")
```

---

mcmc.target.seq      *Find MCMC Target Sequence*

---

**Description**

mcmc.target.seq uses MCMC to find a configuration of DNA positions to get as close as possible to a given tree.

boot.samples.idx uses bootstraps over indices into a DNA matrix.

lookup.samples goes from an index representation of a configuration of DNA to the actual DNABin format.

convert.table.to.idx converts a table of counts for positions 1..n into a list of indices corresponding to positions (i.e. goes from the tabled form to a vector whose tabling matches the input).

**Usage**

```
mcmc.target.seq(data, x, F, n)

boot.samples.idx(data, B = 100, block = 1)

lookup.samples(data, idxs)

convert.table.to.idx(T)
```

**Arguments**

data	A DNA matrix in DNABin format.
x	A tree of class 'phylo' to estimate.
F	A tree estimation function, accepting a DNA matrix in DNABin format and returning a tree of class 'phylo.'



n	The number of MCMC iterations to perform
B	The number of bootstrap replicates.
block	The block size to use during bootstrapping.
idxs	A list of numeric vectors of indices to use for lookup.
T	A table or table-like vector to convert.

### Details

`mcmc.target.seq` performs an MCMC with simulated annealing to locate a configuration of DNA positions from the original matrix that gets as close as possible to a target tree. Propositions for the MCMC replacing one character with another uniformly at random.

The remaining functions are intended to be used as support functions.

### Value

`mcmc.target.seq` returns a list of 4 elements: a numeric vector of counts of each position in the original matrix, the best estimated tree, the best distance from the estimated tree to the target tree, and a numeric vector of the distances for every iteration of the simulation.

`boot.samples.idx`s returns a numeric vector representing the bootstrapped indices.

`lookup.samples` returns a list of objects of class DNABin corresponding to the DNA sequences generated from indices into the original DNA matrix.

`convert.table.to.idx` returns a numeric vector of indices based on the table counts.

### Author(s)

John Chakerian <chakj@stanford.edu>

### References

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Hierarchical Clustering Trees. arXiv:1006.1015v1.

### See Also

[dist.multiPhylo](#) [orthant.boundary.tree](#)

### Examples

```
data(woodmouse)
otree <- root(fastme.ols(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
breps = 200

trees <- boot.phylo(otree, woodmouse, B=breps, function(x)
  root(fastme.ols(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)

combined.trees <- c(list(otree), trees$trees)
```

```
binning <- bin.multiPhylo(combined.trees)

tree.a <- combined.trees[[match(1, binning)]]
i = 2
max.bin = max(binning)
tree.b <- combined.trees[[match(2, binning)]]

while(length(distinct.edges(tree.a,tree.b)) > 1 && i < max.bin)
{
  i = i + 1
  tree.b = combined.trees[[match(i, binning)]]
}

bdy.tree <- orthant.boundary.tree(tree.a, tree.b)

f.est <- function(x) root(nj(dist.dna(x)), "No305", resolve.root=TRUE)

res <- mcmc.target.seq(woodmouse, bdy.tree, f.est, 1000)

par(mfrow=c(2,1))
plot(res$tree)
plot(res$vals)
```

---

orthant.boundary.tree *Orthant Boundary Tree*

---

### Description

Produces a degenerate tree on the boundary between trees that differ by one split.

### Usage

```
orthant.boundary.tree(x,y)
```

### Arguments

x	The tree in the first orthant.
y	The tree in the second orthant.

### Details

The tree found is the tree on the boundary between the two orthants such that it is on the straight line connecting the two trees when one orthant is thought of as being the (-,+) quadrant and the second orthant as being the (+,+) quadrant, where the (0,y) line is the particular boundary in question.

### Value

Returns an object of class 'phylo' representing the boundary tree.

**Author(s)**

John Chakerian <chakj@stanford.edu>

**References**

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

**See Also**

[mcmc.target.seq](#)

**Examples**

```
data(woodmouse)
otree <- root(fastme.ols(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
brep = 200

trees <- boot.phylo(otree, woodmouse, B=brep, function(x)
  root(fastme.ols(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)

combined.trees <- c(list(otree), trees$trees)

binning <- bin.multiPhylo(combined.trees)

tree.a <- combined.trees[[match(1, binning)]]
i = 2
max.bin = max(binning)
tree.b <- combined.trees[[match(2, binning)]]

while(length(distinct.edges(tree.a, tree.b)) > 1 && i < max.bin)
{
  i = i + 1
  tree.b = combined.trees[[match(i, binning)]]
}

plot(orthant.boundary.tree(tree.a, tree.b))
```

**Description**

A family of functions for determining and plotting the differences between two trees. phylo.diff plots two trees side by side, highlighting edges unique to each tree in red. distinct.edges finds the edges present in the first argument not in the second.

`edge.from.split` locates the edge id from a given split.  
`get.bipartition` gets the bipartition of tips formed by a single edge.  
`partition.leaves` returns the set of all bipartitions from all edges.

### Usage

```
phylo.diff(x, y, ...)  
  
distinct.edges(x, y)  
  
edge.from.split(x, split)  
  
get.bipartition(x, e)  
  
partition.leaves(x)
```

### Arguments

<code>x</code>	The first (or only) tree.
<code>y</code>	The second tree, for the functions that accept two trees.
<code>split</code>	A list of bipartitions, probably from <code>partition.leaves</code> .
<code>e</code>	An edge for a particular tree, given as an id.
<code>...</code>	Additional arguments to pass to the <code>plot.phylo</code> function.

### Details

`phylo.diff` uses the ape tree plotting function. The other functions are mostly meant as support functions.

### Value

`phylo.diff` returns invisible.  
`distinct.edges` returns a numeric vector of edge ids for the first tree.  
`edge.from.split` returns an edge id for a particular tree corresponding to a given bipartition and NA if none such edge exists.  
`get.bipartition` returns a character vector of the tips below that edge in the given tree.  
`partition.leaves` returns a list of partitions (themselves character vectors) of the given tree.

### Author(s)

John Chakerian <chakj@stanford.edu>

### References

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

**See Also**[dist.multiPhylo](#)**Examples**

```

data(woodmouse)
otree <- root(fastme.ols(dist.dna(woodmouse)), "No305", resolve.root=TRUE)
breps = 10

trees <- boot.phylo(otree, woodmouse, B=breps, function(x)
  root(fastme.ols(dist.dna(x)), "No305", resolve.root=TRUE), trees=TRUE)

combined.trees <- c(list(otree), trees$trees)

binning <- bin.multiPhylo(combined.trees)

phylo.diff(combined.trees[[match(1, binning)]], combined.trees[[match(2, binning)]])

```

---

position.leverage      *Position Leverage*

---

**Description**

Provides a rough heuristic for determining the degree to which each position in the DNA matrix affects the resulting tree.

**Usage**

```
position.leverage(data, F, to = NULL, rep = 50, by = 1)
```

**Arguments**

data	A DNA matrix in DNAbin format.
F	A tree estimation function, accepting a DNA matrix of class DNAbin and returning a tree of class phylo
to	The tree with which distances are measured in respect to, or NULL to indicate the tree estimated by F for the starting DNA matrix.
rep	The number of times to replicate the position in question.
by	The function will perform the calculation on every by-th position (that is, on seq(1,N,by) )

**Details**

This function takes a DNA matrix and, for every *by*-th position, replicates that position *rep* times, randomly removing *rep* other positions to keep all sequences the same length other positions to keep all sequences the same length. For each new DNA matrix created in this way, *F* is used to estimate the corresponding tree, and the distance to tree *to* is computed and stored. This distance can be thought of as somewhat analogous to the leverage of that position.

**Value**

Returns a numeric vector of distances from tree *to* for each position sampled.

**Author(s)**

John Chakerian <chakj@stanford.edu>

**References**

Chakerian, J. and Holmes, S. P. Computational Tools for Evaluating Phylogenetic and Heirarchical Clustering Trees. arXiv:1006.1015v1.

**See Also**

[dist.multiPhylo](#)

**Examples**

```
data(woodmouse)
f.est <- function(x) root(nj(dist.dna(x)), "No305", resolve.root=TRUE)
position.leverage(woodmouse, f.est, by=10)
```

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